

# PARTIAL CLONING AND APICAL MEMBRANE LOCALIZATION OF AN ANION EXCHANGER (AE-2) IN SHARK (*SQUALUS ACANTHIAS*) RECTAL GLAND TUBULES: PRELIMINARY STUDIES

Albert A. George<sup>1,2</sup>, Rudiger W. Lechrich<sup>3</sup>, Stephen G. Aller<sup>1,4</sup>, and John N. Forrest, Jr.<sup>1</sup>

<sup>1</sup>Department of Medicine, Yale University School of Medicine, New Haven, CT 06510

<sup>2</sup>Savannah State University, Savannah, GA 31404

<sup>3</sup>Humboldt University, Berlin, Germany D-10099

<sup>4</sup>University of New Haven, West Haven, CT 06516

Electroneutral anion exchangers are involved in the regulation of cell volume, intracellular pH and chloride concentration. The first plasma membrane anion exchanger (AE-1) to be identified, cloned and sequenced, was the erythroid band 3, a transporter which mediates one to one exchange of HCO<sub>3</sub> for Cl<sup>-</sup>. By Northern blot analysis and immunohistochemical localization, a Band 3 related protein (AE-2) was identified in a non-erythroid tissue, the mammalian kidney collecting duct (Alper et al., J. Biol. Chem. 263:17092-99, 1989). AE-2 cDNAs have been characterized in mouse and human kidney, rat stomach, (Alper, Ann. Rev. Physiol. 53:549-564, 1991) and rabbit ileal enterocytes (Chow et al., Am. J. Physiol. 263:G345-G352, 1992). Highest expression of AE-2 is seen in several well differentiated cell types including choroid plexus, acid secreting parietal cells of the mammalian stomach, osteoclasts, and renal medulla. *Xenopus* oocytes lack endogenous AE-2 but have an endogenous sodium-hydrogen exchanger (NHE). When AE-2 is overexpressed in *Xenopus* oocytes, tight coupling of endogenous NHE and AE-2 occurs permitting regulatory volume increase (RVI) through net transport of NaCl (Jiang et al., Amer. J. Physiol. 272:C191-202, 1997).

We have been using PCR based strategies to clone membrane proteins involved in regulation of NaCl secretion in the shark (*Squalus acanthias*) rectal gland (SRG). Using highly degenerate primers to 7 transmembrane G protein coupled receptors, we unexpectedly amplified a 555 bp product from SRG which had 88% homology to human AE-2. In preliminary immunohistochemical studies using an antibody to AE-2 (provided by Dr. Seth Alper, Beth Israel Hospital, Boston, MA) we detected apical membrane localization of the AE-2 protein in SRG tubules by confocal microscopy (figure 1). Immunofluorescence was abolished by preincubation with the antigen peptide used to raise the antibody (data not shown). We have also used a <sup>32</sup>P-labeled AE-2 shark specific probe to carry out library screening of a SRG cDNA library and have isolated 25 positive plaques which will be sequenced in pursuit of the full length gene product.



Figure 1. Apical localization of shark AE-2 in rectal gland cells by confocal immunohistochemistry.

In human tissues, AE-2 has been localized by immunohistochemical techniques to apical membranes of small intrahepatic bile ducts and gall bladder (Scoazec et al., J. Hepatol. 26:543-53, 1997). We report here the first identification and partial cloning of an AE-2 exchanger in a marine species. The protein appears to be localized primarily to apical membrane domains of rectal gland tubules. The function of AE-2 in the shark rectal gland tubule is presently under study.

This work was supported by NIH grants DK 34208 and NIEHS P30-ES 3828 (Center for Membrane Toxicology Studies), and a Grant in Aid from the American Heart Association, Maine Affiliate.