TWO TYPES OF Ca²⁺ CHANNELS IN CARDIAC MYOCYTES FROM <u>SQUALUS</u> ACANTHIAS

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Two types of Ca^{2+} channels have been described in cardiac myocytes, which are characterized based on their threshold of activation and speed of inactivation (e.g., Bean, J. Gen. Physiol., 86:1, 1985; Mitra and Morad, PNAS 83: 5340, 1986). The low threshold cardiac t-channel has been described mostly in only mammalian atrial and ventricular myocytes, and its existence in other species has not been widely investigated. In this report, we have investigated whether t-type channels might exist in ventricular myocytes from the dogfish <u>Squalus acanthias</u>, and present evidence below for the existence of both t- and 1-type Ca^{2+} channels in this species.

Single myocytes were isolated enzymatically from shark heart (<u>Squalus</u> <u>acanthius</u>) as described elsewhere (Sorbera et al., this Bulletin) and were studied using the whole cell voltage clamp technique. In TTX-treated (10 uM) myocytes, depolarizing pulses frlom a holding potential of -80 mV, activated first a rapidly inactivating inward current which was maximally activated at -30 mV, which had a voltage dependence and kinetics of activation similar to the t-channel measured in mammalian cardiac myocytes. Depolarization to potentials positive to -20 mV also activated the slower inactivating high threshold Ca^{2+} channel which attained its maximum value at 0 mV.

The two types of inward current showed a differential sensitivity to holding potential as is shown in Fig. 1. When the membrane was held at -80 mV, depolarizing pulses activated both types of inward current (Fig. 1A. However, if a conditioning pre-pulse was applied to -40 mV, only the slowly inactivating current could be activated (Fig. 1B). Subtraction of the currents recorded in Fig. 1A and Fig. 1B yielded a rapidly inactivating inward current alone (Fig. 1C). We further investigated the voltagedependent inactivation of the rapidly inactivating inward current inward current by varying the duration of the conditioning pulse to -40 mV (Fig. 1D). Substantially all of this inward current was abolished following a 500 ms conditioning pre-pulse, the time constant for this process being 118 ms (Fig. 1E).

Further evidence for the existence of two types of Ca^{2+} channel was obtained from the differential sensitivity of these channels to the inorganic Ca^{2+} channel blockers Cd^{2+} and Ni^{2+} . 100 uM of Cd^{2+} blocked all the slowly inactivating inward current, leaving a rapidly inactivating Ca^{2+} current whose voltage range was identical to the difference current shown in Fig. 1C (Fig. 2A). On the other hand, 500 uM of Ni²⁺ preferentially blocked the rapidly inactivating component.

These data are consistent with the existence in shark heart of both land t-type Ca^{2+} channels. The striking finding, however, is the relative size of the t-current compared to the l-current. As can be seen from Fig. 1, the peak currents are approximately equal. This is in stark contrast to the data obtained in mammalian preparations where the t-channel represents at most 30% of the total peak calcium current (Mitra and Morad). This suggests that the t-channel may play a more important role in the shark myocyte than in the mammal.

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