

# MECHANISMS OF $\text{NH}_4^+$ TRANSPORT ACROSS THE DOGFISH PUP (SQUALUS ACANTHIAS) GILL

David H. Evans and Steven D. Robbins

Department of Zoology, University of Florida, Gainesville, FL 32611

Recent studies in our laboratory have demonstrated that ammonia can traverse the teleost and hagfish gill as  $\text{NH}_4^+$ , both by diffusion and via basolateral  $\text{Na}^+/\text{NH}_4^+$  exchange (Goldstein et al., J. Exp. Zool. 219, 395-397, 1982; Claiborne et al., J. Exp. Biol. 96, 431-434, 1982; Evans & Robbins, Bull. MDIBL 84, 52-53, 1984). The present study was undertaken to determine the modes of  $\text{NH}_4^+$  transport across the shark gill epithelium, and the relative permeability to  $\text{NH}_4^+$  of that epithelium.

Heads of dogfish pups were perfused as described previously (Evans & Claiborne, J. Exp. Biol. 105, 363-371, 1983), except that perfusion and irrigation flows were set at 650  $\mu\text{l}/\text{min}$  and 10  $\text{ml}/\text{min}$  respectively, and the perfusate did not contain epinephrine and was aerated with 1%  $\text{CO}_2$  in air. The head was encased in a plastic chamber cooled to 10-15°C by iced tap water circulating through a water jacket, and the irrigation fluid was maintained in the same temperature range by a cooling plate. To test for a role of basolateral  $\text{Na}^+/\text{NH}_4^+$  exchange in baseline total ammonia ( $\text{NH}_3 + \text{NH}_4^+$  - denoted as  $T_{\text{amm}}$ ) efflux, the head was perfused for 20 minutes with elasmobranch Ringer's solution (adjusted to pH ca. 7.8 by the addition of 20 mM  $\text{NaHCO}_3$ ) containing ca. 500  $\mu\text{M}$   $T_{\text{amm}}$ , then for a second 20 minute period with the same perfusate containing  $10^{-4}$  M ouabain. In a second series of experiments, the effect of specific increases in perfusate  $\text{NH}_4^+$  concentration on  $T_{\text{amm}}$  efflux was examined by perfusing the head during the first period with perfusate whose  $T_{\text{amm}}$  concentration had been adjusted to ca. 1 mM (pH ca. 7.8) and during the second period with perfusate whose  $T_{\text{amm}}$  concentration had been adjusted to ca. 9 mM and pH lowered to ca. 6.8 by lowering the bicarbonate concentration to 2 mM, in order to maintain perfusate  $\text{NH}_3$  concentration relatively constant. In these experiments, the head was perfused in a third, 20-minute period with either  $10^{-4}$  M ouabain in the perfusate or  $\text{Na}^+$ -free artificial sea water for the irrigation solution, in order to test for a role of either basolateral  $\text{Na}^+/\text{NH}_4^+$  exchange or apical  $\text{Na}^+/\text{NH}_4^+$  exchange in the  $\text{NH}_4^+$ -stimulated  $T_{\text{amm}}$  efflux. In all experiments, pH and  $T_{\text{amm}}$  of perfusate were monitored at T-10 (10 min after the start of the experiment) of a given experimental time period and pH and  $T_{\text{amm}}$  of irrigate were monitored at T-0 and T-20 of each experimental time period. In this way actual  $\text{NH}_3$  and  $\text{NH}_4^+$  gradients were monitored in each experiment. Since subsequent experiments have shown a relatively high  $\text{NH}_3$  permeability of shark gill epithelium (Evans, More, Robbins, unpublished), changes in  $T_{\text{amm}}$  efflux produced by inadvertent changes in the  $\text{NH}_3$  gradient were factored out for each experiment. To test for structural leaks during perfusion, some heads were perfused with perfusate containing 15mg/ml of eriochlorine (Acid blue 9) for a final 20 minute period. T-0 and T-20 samples were monitored spectrophotometrically (632 nm) and compared with standard concentration vs. color curves for the dye.

Addition of  $10^{-4}$  ouabain to the perfusate reduced the  $T_{\text{amm}}$  efflux from 27 to 13 and 35 to 15  $\mu\text{mol} \cdot 100 \cdot \text{g}^{-1} \cdot \text{hr}^{-1}$  from two heads perfused with 730 and 244  $\mu\text{M}$   $T_{\text{amm}}$ , respectively. Ouabain did increase the afferent pressure (proportional to gill resistance) by some 12 and 15 torr, respectively, but other experiments (Evans, More, Robbins, unpublished results) have

demonstrated that, over a similar range of pressures,  $T_{\text{amm}}$  efflux is not correlated with pressure. So it is unlikely that the significant reduction in  $T_{\text{amm}}$  efflux is secondary to the hemodynamic effects of ouabain. Thus, it appears that a significant percentage (ca. 50%) of the "baseline"  $T_{\text{amm}}$  efflux runs through  $\text{Na}^+/\text{NH}_4^+$  exchange via basolateral Na-K-activated ATPase. Similar results have been found with the teleost *Opsanus beta* (Claiborne et al., op. cit., 1982; Evans & More, unpublished). When the perfusate  $\text{NH}_4^+$  concentration was raised from  $1.3 \pm 0.1 \text{ mM}$  (SE) to  $9.1 \pm 0.3 \text{ mM}$  (9) the  $T_{\text{amm}}$  efflux increased significantly with a slope of  $2.2 \pm 0.7 \mu\text{mol} \cdot 100\text{g} \cdot \text{hr}^{-1} \cdot \text{mM}^{-1} \cdot 1^{-1}$ . Subsequent addition of  $10^{-4}$  ouabain to the perfusate in four of these experiments did not alter the  $T_{\text{amm}}$  efflux ( $+ 7.8 \pm 8.1 \mu\text{mol} \cdot 100\text{g}^{-1} \cdot \text{hr}^{-1}$ ), so it appears that enhanced  $T_{\text{amm}}$  efflux does not run through basolateral  $\text{Na}^+/\text{NH}_4^+$  exchange. In these ouabain experiments, the increase in afferent pressure was only  $6 \pm 2$  torr. Use of  $\text{Na}^+$ -free artificial seawater irrigation during the third period in five experiments actually stimulated the efflux by  $32 \pm 16 \mu\text{mol} \cdot 100\text{g}^{-1} \cdot \text{hr}^{-1}$ , rather than inhibit it as one would expect if apical  $\text{Na}^+/\text{NH}_4^+$  exchange played a major role in  $\text{NH}_4^+$ -stimulated  $T_{\text{amm}}$  efflux. Perfusion with Acid Blue 9 demonstrated that the structural leak after some 90 minutes of perfusion was only  $1.8 \pm 0.7\%$  (7), which could account for an apparent  $T_{\text{amm}}$  efflux of only some  $0.5 \mu\text{mol} \cdot 100\text{g}^{-1} \cdot \text{hr}^{-1}$ , approximately 1% of that actually measured in these experiments. Therefore, structural leaks obviously cannot account for the  $T_{\text{amm}}$  effluxes from the perfused pup head. Thus, the slope of the  $\text{NH}_4^+$ -stimulated  $T_{\text{amm}}$  efflux is proportional to the gill epithelium's permeability to that cation.

If we assume that the functional branchial surface area of the pup is approximated by the structural surface area of the adult dogfish ( $3.7 \text{ cm}^2/\text{g}$ ; Hughes & Wright, Z. Zellforsch mikrosk. Anat. 104, 478-493, 1970), we can calculate that the apparent  $P_{\text{NH}_4^+}$  is  $1.7 \times 10^{-6} \text{ cm} \cdot \text{s}^{-1}$ , somewhat less than (but of the same order as) that described for the turtle bladder ( $4.5\text{--}4.9 \times 10^{-6} \text{ cm} \cdot \text{s}^{-1}$ ; Arruda et al., Am. J. Physiol. 246, F635-F647, 1984; Schwartz & Tripolone, Am. J. Physiol. 245, F210-F216, 1983). Since it is likely that the functional surface area of the branchial epithelium is somewhat less than the structural surface area (and therefore the apparent  $P_{\text{NH}_4^+}$  is somewhat underestimated) it is clear that the  $P_{\text{NH}_4^+}$  of the shark pup gill is similar to that described for a model tight epithelium like the turtle bladder. The relatively low  $P_{\text{NH}_4^+}$  of the shark gill does correlate with its relatively low permeability to  $\text{Na}^+$  (Evans, in Osmotic and Ionic Regulation in Animals, ed. by G. M. O. Malooy, Academic Press, pp. 305-390, 1979). Subsequent experiments in our laboratory in Gainesville have determined that the apparent  $P_{\text{NH}_3}$  of the shark pup gill is  $4.2 \times 10^{-4} \text{ cm} \cdot \text{s}^{-1}$ , somewhat higher than that described for the turtle bladder ( $0.7$  and  $2.6 \times 10^{-4} \text{ cm} \cdot \text{s}^{-1}$ ; Arruda et al., 1984, op. cit., and Schwartz & Tripolone, 1983, op. cit., respectively).

In summary, our data indicate that the branchial epithelium of the dogfish shark is approximately 250 times as permeable to  $\text{NH}_3$  as to  $\text{NH}_4^+$ . However, basolateral  $\text{Na}^+/\text{NH}_4^+$  exchange can account for approximately 50% of the total ammonia transport, and the fact that the pH of shark blood is 2 pH units below the pK for ammonia dictates that, of the remaining 50% of ammonia efflux, 14% is via ionic diffusion and 35% is via non-ionic diffusion. Supported by NSF PCM 8302627.