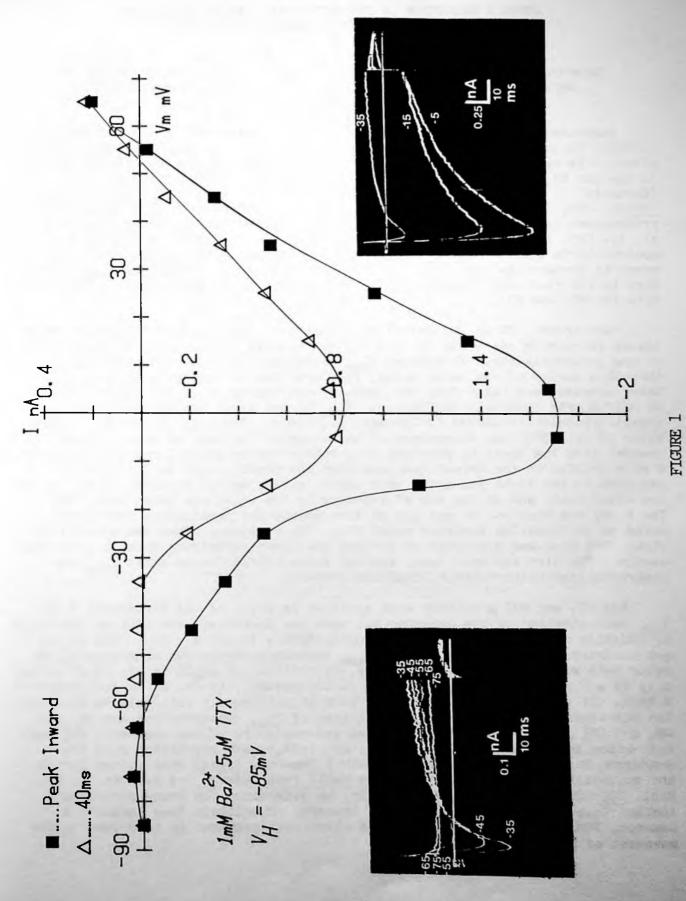
EVIDENCE FOR TWO POPULATIONS OF CA²⁺ CHANNEL IN VENTRICULAR CELLS OF SQUALUS ACANTHIAS

R. Mitra & M. Morad, Department of Physiology, University of Pennsylvania, School of Medicine, Philadelphia, PA 19104

In the previous communication (MDIBL Bulletin, 1985) we have characterized the membrane currents in isolated ventricular myocytes of shark heart using the giga-seal technique (Hamill et al., Pfulgers Arch. 391:85, 1981). In these cells we were able to identify a fast TTX-sensitive Na channel, and an inwardly rectifying K channel, in addition to the Ca channel. However, the Ca channel could not be easily classified as a unitary transport system.

Analysis of inactivation kinetics of Ca²⁺ current yielded a fast (10 ms) and a slower (86 ms) time constant suggesting at least a 2 step process for the inactivation of this current. Examination of the voltage-dependence of Ca²⁺ current yielded an I-V relation not consistent with one population of channels. In TTX (5 uM) containing solutions with holding potentials negative to -80 mV, a TTX insensitive fast inward current could be demonstrated 1. This current activated from -60 to -50 mV and was blocked by Co²⁺ but not by Cd²⁺. At more positive holding potentials (-50 to -40 mV), the conventional Ca²⁺ current described earlier in this preparation and well studied in other species could be recorded. Figure 1 illustrates an attempt to separate the two populations of Ca²⁺ channels based upon their time and voltage dependence. Note that while holding at -85 mV, the peak inward current in the presence of TTX and Ba²⁺ has a distinct shoulder at about -30 mV where the conventional Ca²⁺ current is generally activated. The I-V relation measured 40 ms into the clamp step shows activation voltage for the channel more similar to the conventional Ca²⁺ current. The insets show that the current activating between -60 and -30 mV has faster kinetics than the current activated positive to -35 mV.

Our results suggest the existence of two distinct populations of Ca channels with different voltage and pharmacological sensitivities in isolated shark ventricular myocytes.



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