

Table III
 Thickness of the Cornea Measured with the Specular Microscope for an Elasmobranch and a Teleost
 and the Effect of Low Temperature

| | Control | | | Depth of Anterior Chamber | Low Temperature* | | Epithelium |
|--|-------------------------|------------------------------|----------------------|---------------------------|-------------------------|---------------------------|------------|
| | Total Thickness Central | Periferal | Epithelium | | Total Thickness Central | Periferal | |
| Spiny Dogfish <i>(Squalus acanthias)</i> | | | | | | | |
| Means | 161 | 184 | 66 | 342 | 159 | 155 | 67 |
| ±SEM | 9 | 17 | 6 | 102 | 11 | 13 | 3 |
| N | 14 | 9 | 6 | 6 | 7 | 7 | 4 |
| <hr/> | | | | | | | |
| Longhorn Sculpin <i>(Myoxcephalus Octodecimspinosus)</i> | Epithelium | Intermediate Layer of Stroma | Descemet-Endothelium | Total Thickness of Cornea | | Depth of Anterior Chamber | |
| | | | | Control | Cold * | | |
| Means | 14 | 124 | 213 | 241 | 820 | 725 | |
| ±SEM | 3 | 7 | 16 | 12 | 90 | 129 | |
| N | 4 | 7 | 4 | 7 | 7 | 6 | |

* Eye kept overnight at 2° C covered with Ringer.

In the teleosts the images obtained were 5. They correspond to the layers of reflexion of the images in the complex cornea of these fish. The first two images corresponded to the anterior and posterior portion of the epithelium, the third to the anterior or dermal portion of the stroma, and the fourth and fifth to the outer and inner limits of the Descemetendothelial zone. The actual thickness of the regions and the effect of low temperature is shown in Table 3. It is evident that the cornea of teleosts swells and increases its thickness in the cold.

In summary, the ancient cornea of the shark, possesses the ability to remain at normal hydration and thickness in adverse metabolic conditions, and at the same time, no significant electrical potential difference is found arising from the epithelial layers. On the other hand, in the more recent teleosts, the cornea swells in the cold and shows the consistent presence of an electrical potential difference arising from the epithelium, similar to amphibians and mammalians. Specular microscope measurements of corneal thickness *in vivo* and in mounted corneas of sharks and sculpins is reported.

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Plasma And Urine Proteins In The Agglomerular Teleost, *Lophius americanus*

R. Galaske, State University of N.Y. at Buffalo and Medizinische Hochschule, Hannover

Bieter, using the less sensitive methods of his time, found no protein in the urine of *Lophius* (*J. Pharm. and Exper. Therap.* 43:406, 1931). His finding that injected hemoglobin appeared in the urine of glomerular but not of

agglomerular fish led to the conclusion that most urinary protein was of glomerular origin.

A more valid examination of the renal handling of plasma proteins in agglomerular fish has been made possible by the recent development of micro techniques of polyacrylamide gel electrophoresis. The appearance of any plasma protein in the urine of an agglomerular fish is of interest with respect to its mode of access, secretory or otherwise. Moreover, since filtration and tubular reabsorption of low molecular weight proteins (LMW) are important processes in their catabolism in a filtering kidney we may question if accumulation in plasma or alternate pathways of disposal for LMW proteins have evolved with the loss of glomerular function.

In this report data are presented for the renal clearance of plasma proteins by *Lophius* and for the clearance of an inert macromolecule (PVP) injected intravenously.

Clearance experiments were performed in male fish weighing from 2000 to 3000 g. Fish were placed ventral side up with head and gills submerged. Both ureters were catheterized with PE tubing through a ventral abdominal incision. Blood samples were taken from the caudal vein. Total protein concentration in plasma and urine were measured by the Lowry method. Micro-polyacrylamide electrophoresis with a continuous gradient from 2 to 40% was performed on diluted plasma and unconcentrated urine samples. Some urine samples were concentrated by ultrafiltration in dialysis tubing (Union Carbide) and Amicon filters (UM 2). Concentrated urine or unconcentrated plasma was separated by gel filtration on Sephadex G 100, G 75 and G 50 columns. Reconcentrated eluted fractions were again separated by gradient gel electrophoresis. One fish was injected with PVP (MW 40,000) and renal clearance and plasma disappearance curve of macromolecules were determined.

PLASMA (*Lophius americanus*)

l:20
0.5 μ l

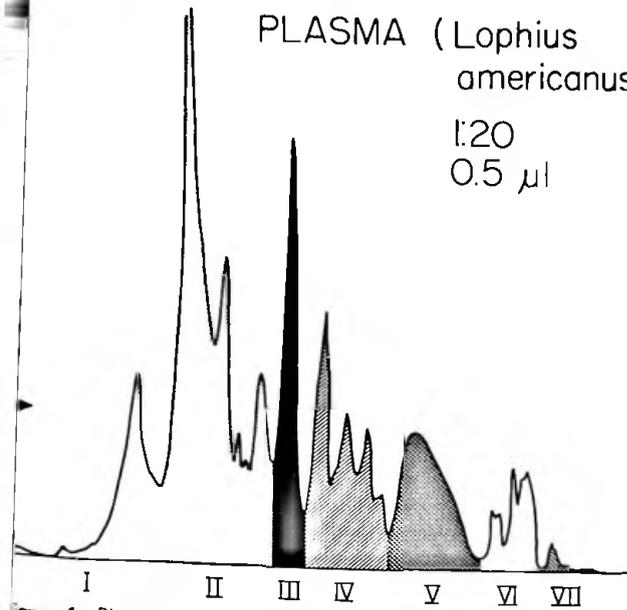


Figure 1 Plasma protein pattern in gradient-gel-electrophoresis (0.5 μ l of 20x diluted fish plasma). The direction of migration is from left to right. Roman numerals indicate different fractions.

URINE (*Lophius americanus*)
50x conc.
0.5 μ l

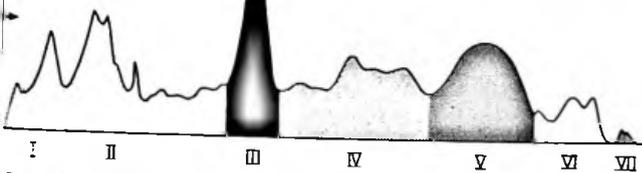


Figure 2 Urinary protein pattern in gradient-gel-electrophoresis (0.5 μ l of 50x concentrated fish urine). Fraction numbers (Roman numerals) correspond to fraction numbers in Figure 1.

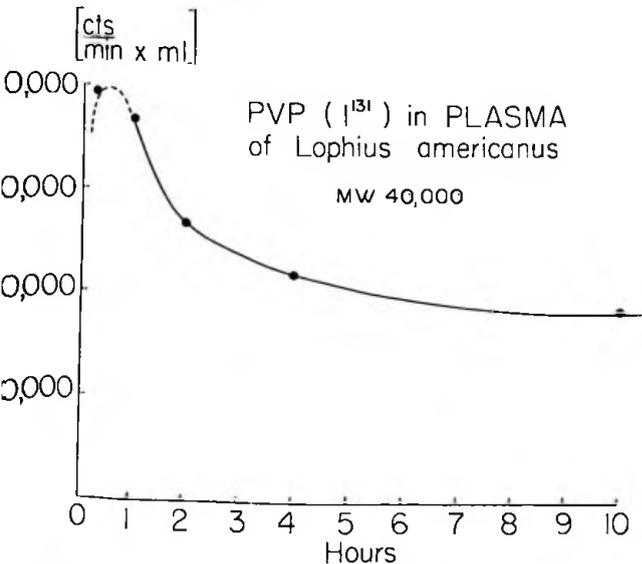


Figure 3 Disappearance curve of PVP from plasma.

Plasma total protein concentration was 2032 ± 92 (SD, $n=5$) mg/100 μ l and urinary protein was 0.72 ± 0.07 ($n=19$ mg/100ml).

Fig. 1 shows the pattern of plasma electrophoresis of 0.5 μ l of 20x diluted plasma. Proteins, 32 in number, were divided into 7 major fractions, according to their separation on Sephadex G100 gel. The MW range is = 100 000(#1), 70 000(#2), 55 000(#3), 45 000(#4), 33 000(#5), 18-25 000(#6) and 11-15 000(#7). There is no redominant plasma protein band corresponding to albumin in mammalian plasma. The fraction of proteins smaller than 55,000 MW comprises $27.8 \pm 3.4\%$ ($n=5$) in plasma. Fraction #3 (55,000) is $13.4 \pm 1.2\%$ and higher molecular weight proteins are $59 \pm 2.8\%$.

Fig. 2 shows the pattern of urinary proteins in 0.5 μ l of 50x concentrated fish urine. The curve reveals the whole spectrum of plasma proteins, but at a very low concentration (total protein = 0.72 ± 0.07 mg/100ml).

Table 1 shows the renal clearances of these proteins as calculated for one kidney and the renal clearances for PVP in the same kidney and time period. The slope for the disappearance of PVP from plasma in the same animal is plotted in Fig. 3.

Quantification of the different protein groups was performed by integrating the corresponding electrophoretic bands. Results are shown in Table 2. Fraction #3 (MW 55 000) was used as reference for establishing a ratio of molecules smaller than 55,000 (LMW proteins) and larger than 55,000.

Molecules smaller than 55,000 comprise in plasma $27.8 \pm 3.4\%$ ($n=5$) and in urine $59 \pm 4.2\%$ ($n=3$), whereas

Table I

$C_{\text{PROTEIN}} \text{ \& } C_{\text{PVP}} \text{ min}^{-1} \times 100 \text{g BW} \times 1 \text{ Kidney}$

| t-Period | C_{PROTEIN} Kidney | | C_{PVP} Kidney | |
|----------|--------------------------------|----------------------|----------------------------|---------------------|
| | Left | Right | Left | Right |
| U2 | $0.8 \cdot 10^{-6}$ | $0.5 \cdot 10^{-6}$ | $1.8 \cdot 10^{-6}$ | $2.7 \cdot 10^{-6}$ |
| U3 | $0.6 \cdot 10^{-6}$ | $0.35 \cdot 10^{-6}$ | $2.5 \cdot 10^{-6}$ | $2.6 \cdot 10^{-6}$ |
| U4 | $0.18 \cdot 10^{-6}$ | $0.62 \cdot 10^{-6}$ | $2.7 \cdot 10^{-6}$ | $3.0 \cdot 10^{-6}$ |
| U5 | $0.45 \cdot 10^{-6}$ | $0.24 \cdot 10^{-6}$ | $2.1 \cdot 10^{-6}$ | $2.2 \cdot 10^{-6}$ |
| U6 | | | $2.7 \cdot 10^{-6}$ | $2.3 \cdot 10^{-6}$ |

Renal clearances of total protein (C_{protein}) and PVP (C_{PVP}) (i.v.; M.W. 40,000) for 2 kidneys of the same animal. (L3).

Table II

Fractions of Plasma Proteins
in Gradient Gel Electrophoresis

| | <55,000 | =55,000 | >55,000 |
|--------|-------------------------------|-------------------------------|-------------------------------|
| Urine | $59.0 \pm 4.2\%$ ($n=3$) | $19.0 \pm 1.5\%$ ($n=3$) | $22.0 \pm 5.0\%$ ($n=3$) |
| Plasma | 27.8 ± 3.4 ($n=5$) | 13.2 ± 2.8 ($n=5$) | 59.0 ± 2.8 ($n=5$) |

Percent values of proteins with higher and lower MW than 55,000. The fraction with MW 55,000 is identical to fraction III in Figure 1 and Figure 2.

the fraction of high molecular weight proteins (#1) is $22 \pm 5\%$ (n3) in urine and $59 \pm 2.8\%$ (n5) in plasma.

Although the disappearance curve of PVP from *Lophius*' plasma (Fig. 3) follows a time course similar to that in mammalian species the renal clearance of this molecule is incomparably less. In the rat, for example, clearance of PVP (MW 40,000) is 10,000 x the value calculated here for *Lophius*. The extrarenal disposal of this macromolecule is therefore of considerable interest.

When relative clearances of PVP and protein are compared within species another striking difference is found. In *Lophius* (Table 1) the clearance of PVP is 3 or 4 x tht of total plasma protein, in the rat it is about 1000 x. This discrepancy may be explained by the relatively small molecular weight of fish plasma proteins and the large fraction of LMW proteins in the plasma of *Lophius*.

These preliminary data allow no more than inference regarding the mode of access of plasma proteins to the urine in this aglomerular species. The fact that all plasma proteins find urinary representation speaks against secretion since it seems unlikely that secretory mechanisms for every protein would be evolved. Also LMW proteins predominate in urine whereas known secretory proteins (IgA, uromucoid) are of high molecular weight. It is possible that the very small rate of urinary protein excretion (12 g/hr/kidney) represents leakage of plasma proteins through intercellular spaces. The bulk flow of urine across these epithelial structures may assist the movement of proteins. The predominance of LMW proteins in the urine of *Lophius* is consistent with this process of proteinuria.

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Metabolism of the Standing Dead Plant Community in Several Maine Salt Marshes

John L. Gallagher and William J. Pfeiffer. The University of Georgia Marine Institute, Sapelo Island, Georgia

The salt marsh, like many natural grasslands, is a detritus based ecosystem. Relatively few grazers are present and most of the photosynthate enters the food web via microbial degradation. Microbes are active in southeastern United States marshes throughout the year, but in Maine a strong seasonality in plant decay would be expected.

Aerial and aquatic respiration of the dead plant communities and dissolved organic carbon (DOC) release were measured in stands of *Spartina alterniflora* Loisel., *Spartina patens* (Ait.) Muhl. and *Juncus gerardi* Loisel. Samples were collected from marshes at Northeast Creek and Hog Bay in Hancock County. Aquatic respiration of the community associated with the standing dead plants was measured as oxygen consumption by the Winkler method and aerial respiration was quantified as CO₂ release with an infrared gas analyzer. DOC discharge was measured by the method described by Gallagher *et al* (*Estuar. Coast Mar. Sci.*, *in press*).

Aquatic respiration rates of the three dead plant communities were similar in April when the temperature

was 1°C. They were 14.9(\pm 0.6), 12.6(\pm 1.0), 15.4(\pm 0.5) g C hr⁻¹ gram dry weight⁻¹ (\pm SE) for the *alterniflora*, *S. patens* and *J. gerardi* respectively. Over the wide temperature range used to compare the Maine (ambient 1°C) and Georgia (ambient 25°C) communities the Q₁₀ was 2.75. Aerial respiration rates in April were slightly lower than those measured under aquatic conditions. In May the aerial Q₁₀ for the *S. alterniflora* community averaged 3.89 over the temperature range 2-18°C, while that for *S. patens* was 2.77. These values are higher than similar values for Georgia and indicate the saprophages in Maine communities are adapted to take advantage of short periods of warm weather. Carbon dispersion from the dead plant communities by DOC leaching was more than 4 times that of respiration rate. These compounds released from the dead plant communities are readily absorbed by planktonic heterotrophs. Chemical analyses of the dead plant communities for sugars, starch and crude protein are not complete, but similar studies in Georgia have shown a strong correlation between respiration rate and protein content of the dead plants.

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Free Amino Acids In Tissues Of The Skate, *Raja erinacea*: Regulation Of Concentrations And Transport During Adaptation To A Dilute Sea Water Environment.

Leon Goldstein, Thomas A. Boyd, Anne E. McElroy, Chung-Ja Cha and Roy P. Forster Brown University, at Dartmouth College

Study of intracellular osmoregulatory processes is difficult in higher vertebrates because these animals do not survive large changes in osmolarity of body fluids. However, certain fish (euryhaline elasmobranchs) are suited ideally for this type of study, for they tolerate relatively large alterations in body fluid osmolarity. Previous studies (*Forster and Goldstein, Am. J. Physiol. in press*) have shown that amino acids play a major role in intracellular osmoregulation in the skate *Raja erinacea*. Total free amino acids in skeletal muscle and RBC of skates kept in seawater (SW) comprise more than 1 percent of the osmotically active solutes, and this concentration is reduced significantly when the fish are adapted to half-strength sea-water (1/2 SW). In the present study, we measured the levels of individual amino acids in RBC, wing muscle and heart of skates maintained in SW and 1/2 SW.

Blood was drawn from a caudal vessel. The skate was pithed and a piece of wing muscle and heart quickly removed and immediately freeze-clamped with aluminum blocks precooled in liquid nitrogen. Blood was separated into packed RBC and plasma and the latter deproteinized with 10 percent trichloroacetic acid (TCA). The TCA was removed with ether. Frozen, packed RBC, wing muscle and heart were powdered separately in mortar and surrounded with dry ice and extracted with 90 percent sulfosalicylic acid. The tissue and plasma extracts were dried under vacuum, over conc. H₂SO₄, at 40°C. The dried residues were dissolved in appropriate volume